Growth Performance of the Freshwater Mussel, *Unio terminalis delicates* (Lea, 1863) (Mollusca: Bivalvia: Unionidae) in the Gölbaşı Lake, Turkey

Hülya Şereflişan^{1,}* and Mahmut A. Gökçe²

¹Faculty of Marine Sciences and Technology, İskenderun Technical University, 1200 Iskenderun, Hatay, Turkey ²Faculty of Fisheries, Cukurova University, Adana, Turkey

ABSTRACT

In this study, the effects of four different stocking densities (20, 40, 60, 80 individuals/m²) were investigated on growth performance of the *Unio terminalis delicates* in a natural lake (Lake Gölbaşı, Kırıkhan, Hatay, Turkey). Growth in live weight, shell length, width and height were measured monthly for the four different stock groups during 6 months. The highest growth rate was obtained from 40 ind/m²group, while the best condition factor was acquired from the lowest density group (20 ind/m²). The specific growth rate for length and weight were highest in June and July in all groups. On the other hand, plankton biodiversity, chlorophyll a, Mg, NO₂, NO₃, amount of organic matters, NH₃, PO₄, Si, Ca and chemical oxygen demand (COD) were determined as biological and chemical characteristics. At the end of the study, the greatest live weight (5.06 ± 0.34 g) was obtained from 40 ind/m², with 3.34 ± 0.05 cm, 1.36 ± 0.03 cm and 1.91 ± 0.04 cm as length, width and height values, respectively. Therefore, it could be advised that juveniles should be stocked in early spring for *U. terminalis delicates* at a level of 40 ind/m². This results suggest that stocking density is the key factor in mussels culture and important for captive breeding in mussels production.

INTRODUCTION

 ${f F}$ reshwater mussels (Unionidae) are a diverse and conspicuous members of the benthic fauna in fresh waters (Haag and Staton, 2003; Haag, 2013). They play important role in freshwater ecosystem and are economically valuable for their shells (Strayer et al., 2004). The life cycle of unionids is quite different and remarkable (Barnhart, 2006; Şereflişan et al., 2013). In part because of this complex life cycle, mussels are now one of the most imperiled groups of organisms on Earth (Haag, 2013). The bivalve culture has a huge potential in aquaculture sector (McMahon, 1991; Parsons and Dadswell, 1992: Monteforte et al., 1994: Mueller and Patzner, 1996; Morris and Corkum, 1999). Bivalve culture systems are classified into spawning, larval breeding and intensive raising stages. Over the past decade, efforts to propagate and culture unionids have expanded. However, few studies have tested the effects of factors such as temperature, water quality, food type, or food availability on juvenile growth and survival (Barnhart, 2006).

Culture studies carried out on other unionids have

0030-9923/2016/0004-1109 \$ 8.00/0

Copyright 2016 Zoological Society of Pakistan



Article Information Received 6 January 2015 Revised 9 February 2016

Accepted 3 April 2016 Available online 1 June 2016

Authors' Contribution

MAG conceived and designed the study and wrote the article. HS cultured mussels and executed all the experiment work.

Key words

Unio terminalis delicates, cage culture, stocking density, growth performance, water quality parameters

shown differences in growth performances at different locations (Mueller and Patzner, 1996; Paterson and Nell, 1997). These differences were relatively related with environmental factors such as chlorophyll a, organic matter, pH and temperature (Morris and Corkum, 1999). Growth performance varies according to spat stage, planktonic population and density (Chatterji et al., 1984), and water flow rate (Doroudi ve Soutgate, 2000) in the culture area. Spat stage has major affect on the growth performance in culture (Chatterji et al., 1984), because juveniles show an exponential growth when compared to elder individuals (Stanczykowska and Levandowski, 1995). Furthermore, it is claimed that the substrate or depth structure doesn't have as much as effect on the growth and survival rates compared to other factors (Beaty and Neves, 1996; Monteforte and Garcia-Gasca, 1994; Monteforte et al., 1994; Monteforte and Morales-Muliay-Mulia, 2000; Taylor et al., 1998).

In contrast to marine species, freshwater mussels juvenile specimens of many species have bysal thread which is mostly lost in adults (Bogan, 2008). So, cage culture is globally accepted as an efficient method for freshwater mussels. Previous studies on *Crassostrea virginica* (Simmons *et al.*, 1995), *Pinctada margaritifera* (Parsons and Dadswell, 1992), *Placopecten magellanicus* (Parsons and Dadswell, 1992; Southgate and Beer, 1997), *Mytilus edulis* (Karayücel and Karayücel, 2000a), *Pinctada mazatlanica* (Monteforte *et al.*, 1994) and the golden freshwater mussel (*Limnoperna fortunei*)

^{*} Corresponding author: <u>hulyasereflisan@hotmail.com</u> - <u>hovat@mku.edu.tr</u>

(Boltovskoy and Cataldo, 1999) and revealed that stocking density has a significant effect on the growth and survival rate.

The study was conducted in a natural lake which is located in Hatay province, Turkey. The dominant freshwater species of this lake, *Unio terminalis delicates*, has an economical value in southeastern Turkey (Şereflişan, 2014). However, there have been no studies on the environmental requirements, growth and survival rates in cages for *Unio terminalis delicates*. Moreover, there has been no effort to culture this species. Thus, in this study, the effect of different stocking densities in cage culture system on growth performance of *Unio terminalis delicates*, was investigated, to assess the potential to culture *U. terminalis delicates* as an alternative food source.

MATERIALS AND METHODS

Study site

The study was organized between June - November 2001 in Lake Gölbaşı, Kırıkhan, Hatay (36° 30'16"N; 36° 29'42" E). The experiment was carried out in Lake Gölbaşı, Hatay, southern Turkey, between May 2001 and April 2002. Lake Gölbaşı is in the eastern Mediterranean region of Turkey, 50 km north of the city of Antakya. The lake is a natural lake with a surface area of 12 km² at altitude of 80 m (Fig. 1).

Biological materials

Juveniles mussel (1200 Nm) were collected with a scoop net and a hand dredge at a depth of 1-3 m in Lake Gölbaşı. The shell lenght was measuremened with digital caliper. The initial main shell measurements were 6.2±0.08 mm in length, 4.0±0.01 mm in height and 2.5±0.02 mm in width. The average initial weight of the juvenile mussels was 0.64±0.08 g. Juveniles were stocked in cylindrical cages of one cubic m volume (115 cm diameter and 25 cm height). Cages were covered with a metal net (2 cm mesh size), and a polyamide net with 6 mm mesh size covered the inside of the metal net. The experimental design was composed of four stocking density treatments with three replicates each. Four each treatement the stocking density was: A - 20 ind./m2, B -40 ind./m², C - 60 ind./m², and D - 80 ind./m². The juvenile mussels were placed in 12 metal cages at approximately 2 m depth. Sampling and biometry Systematic sampling were conducted according to Hoggarth (1999). Shell length, width, height and live weight measurements were taken monthly for 12 months, while wet meat weight measurements were determined at the end of the study. Measurements were performed on of 20 mussels randomly sampled from each cage. The mantle cavity water of the sampled mussels was drained before weighing. The growth parameters were estimated from the changes in shell length (L), live weight (LW) and wet meat weight (WMW). Mortality was determined by counting the number of empty shells in each cage monthly.

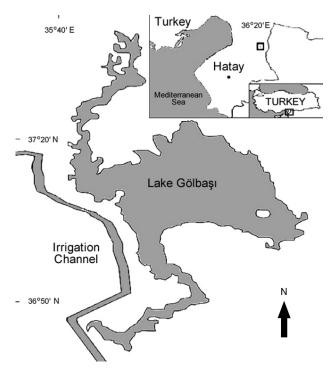


Fig. 1. Map of the Hatay city showing the study site, Gölbaşı Lake, Turkey.

Growth parameters

Growth parameters were determined from the changes in shell length (L), live weight (LW) and wet meat weight (WMW). Mortality was assessed monthly, by counting the number of empty shells in each cage.

Condition factor (Monteforte and Morales-Mulia, 2000) and the specific growth rate (Chatterji *et al.*, 1984) were calculated by using the following equations.

Condition factor = $(TWW/SV) \times 100$

TWW, total wet weight; SV, shell volume

Specific growth rate (%) = $[(lnL_2 - lnL_1) / (T_2 - T_1)] \times 100$

LnL2, ending shell length; LnL1, beginning shell length; T2-T1, the time interval.

Water quality parameters

Oxygen, temperature and pH were measured directly in the field with digital instruments. Oxygen and

temperature were measured by a YSI model 52 oxygen meter and pH by an Orion model 420A pH meter. Nansen-type bottles were used for lake water sampling. Water samples (1 L) were taken to laboratories of Mustafa Kemal University for the analysis chlorophyll a (Ch a), particulate organic matter (POM), ammonia (NH₃), nitrite (NO₂), nitrate (NO₃), calcium (Ca), magnesium (Mg), phosphate (PO₄) and silica (Si). For Ch a and POM analysis, water samples were passed through a 150 µm nylon mesh to remove large particles. The number of particles was determined with a Fluorometrik method (APHA, 1980). The EDTA (Gehrke *et al.*, 1954) method was used to analyze Ca and Mg, whereas a spectrophotometer was employed to analyze NH₃, NO₂, NO₃, PO₄ and Si (APHA, 1971).

Statistical analysis

Analysis of variance (ANOVA) was employed to detect growth differences between the stocking groups with a significance level of 0.05. The results of variance analysis, the lowest standard error and regression coefficients were also evaluated to find the best regression model. Statistics contained in Zar (1999) was used to compare the regression equations of the models. The Excel tool Pack program (Microsoft Office Excel 2013) was used to determine the mean and standard errors for water quality parameters. All statistical analyses were carried out with the SPSS (Statistical Package for the Social Sciences) software.

RESULTS

Shell and tissue growth

The mean weight (W), length (L) and height (H) values of each mussel group are summarized in Figures 2, 3, and 4. All groups showed growth from June to August. The growth in length was highest in groups B (40 ind./m²) and C (60 ind./m²) (Fig. 2). On the other hand, there was no statistical difference on final growth performance between groups B and C, and between groups A and D (p>0.05). Wet meat weight was highest in the stocking group B of 40 ind./m², and wet meat yields of 20 ind./m² and 40 ind./m² were found to be higher than those of the stocking groups of 60 and 80 ind./m² (Fig. 4). A sharp decrease in growth rate was observed from the beginning of the study until August, followed by a more stable period until November (Figs. 2, 3). A negative growth was observed in winter months, followed by quick recovery in spring. The highest condition factor value was obtained from Group A with 13.84±0.69 and the lowest from Group C with 11.21±0.50. There were no statistical differences observed for all groups except Group A (P<0.05).

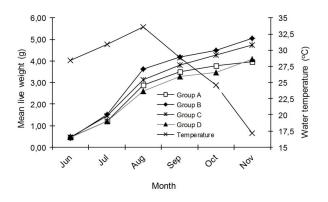


Fig. 2. Monthly comparison of average increase in mean live weight among groups from June to the and of November and relation of growth with temperature. [Stock groups Group A (20 ind./m²), Group B (40 ind./m²), Group C (60 ind./m²), Group D (80 ind./m²)].

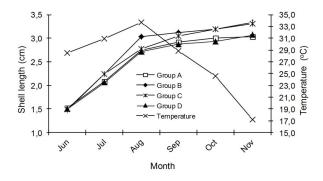


Fig. 3. Comparison of growth in shell length of mussels in relation to temperature on monthly basis (June-November). Stock groups [Group A (20 ind./m²), Group B (40 ind./m²), Group C (60 ind./m²), Group D (80 ind./m²)].

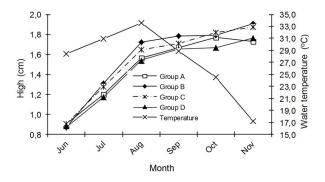
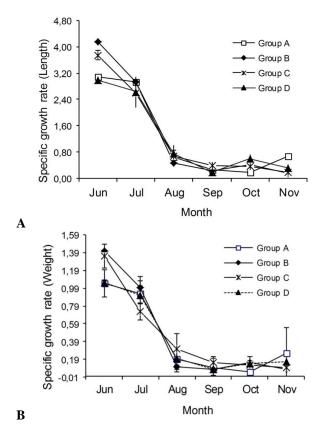
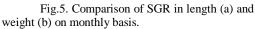


Fig. 4. Comparison of growth in shell high of mussels in relation to temperature on monthly basis (June-November). Stock groups [Group A (20 ind./m²), Group B (40 ind./m²), Group C (60 ind./m²), Group D (80 ind./m²)].

H. ŞEREFLİŞAN AND M.A. GÖKÇE





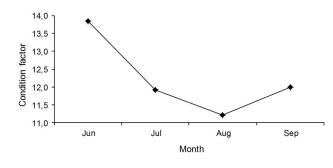


Fig. 6. Variation condition factor according to stock groups at the end of the experiment (November, 2002).

Environmental factors

The lake water temperature ranged from 10.59° C in January to 33.63° C in August. Annual range of oxygen concentration was 4.51 ppm (January) to 9.60 ppm (November). The changes in pH were nearly insignificant during the course of the study. Monthly contents of Ch a, POM, NH₃, NO₂, NO₃, Ca, Mg, PO₄ and Si are shown in Table I. Monthly contents of Ch a, Ca, and Mg varied

Table I	Water quality parameters measured for a duration of 6 months in the Lake Gölbaşı of Turkey (mg I ⁻¹) (Mean ±SE); Ch, Chlorophyll a; Mg, magnesium; NO ₂ , nitrite; POM, particulate organic matters; NH ₃ , ammonia; NO ₃ , nitrate; PO ₄ , phosphate; Si, silica; Ca, calcium; COD, chemical oxygen demand.	neters measured articulate organ	l for a duration nic matters; NH	of 6 months 3, ammonia; N	in the Lake Gi 0 ₃ , nitrate; PC	ilbaşı of Turke 04, phosphate; {	y (mg l ⁻¹) (Mea Si, silica; Ca, ca	n ±SE); Ch, Cl leium; COD, cl	hlorophyll a; N hemical oxyger	fg, magnesium; ı demand.
Months	Cha	Mg	NO2	POM	NH ₃	NO ₃	PO_4	Si	Ca	COD
June	0.53 ± 0.01^{a}	$2,85\pm0,04^{a}$	0,020±0,00°	3,83±0,06 ^b	$0,43\pm0,01^{a}$	$1,20\pm 0,00^{\circ}$	0,03±0,01 ^b	4,45±0,00 ^{cd}	4,42±0,01 ^b	$10,01\pm0,01$
July	$0,25\pm 0,02^{cd}$	$2,81\pm0,02^{a}$	$0,070\pm0,00^{a}$	$0,63\pm0,06^{d}$	$0,18\pm0,00^{\rm b}$	$1,32\pm0,01^{b}$	0,00±0,00 ^{cd}	$3,83\pm0,00^{e}$	$4,58\pm0,01^{a}$	$10,00\pm0,00$
August	$0,22\pm0,01^{e}$	$2,54\pm0,02^{a}$	$0,003\pm0,00^{d}$	1,04±0,01°	0,11±0,01°	$1,14\pm0,00^{\rm d}$	$0,07\pm0,01^{a}$	$11,09\pm0,02^{a}$	4,17±0,02 ^d	$9,43\pm0,17$
September	er 0,24±0,01d	$2,54\pm0,03^{b}$	$0,046\pm 0,00^{b}$	$4,56\pm0,03^{a}$	0,11±0,01°	$1,40\pm0,00^{a}$	$0,01\pm0,00^{c}$	$2,95\pm0,01^{f}$	4,34±0,03°	$8,42\pm0,16$
October	$0,27\pm0,01^{cb}$	$2,54\pm0,03^{b}$	$0,020\pm0,00^{c}$	4,56±0,04ª	$0,17\pm0,01^{b}$	$1,32\pm0,01^{b}$	$0,01\pm0,00^{c}$	4,60±0,00 ^{bc}	4,06±0,04°	6,07±0,06
November	уг 0,29±0,01 ^b	2,54±0,05 ^b	$0,026\pm0,00^{b}$	$4,56\pm0,01^{a}$	$0,19\pm0,00^{d}$	$1,31\pm0,03^{b}$	$0,05\pm0,01^{\text{ba}}$	4,76±0,01 ^b	$4,32\pm 0,04^{\circ}$	5,03±0,04
Letters a. b.	Letters a. b. c. d. e. f and g show statistical differences in vertically (P<0.05).	tistical difference	es in vertically (<i>F</i>	2<0.05).						
	2									

significantly different (P<0.05). The highest Ch a (0.53 \pm 0.01 mg/L) and NO₃ (1.40 \pm 0.00 mg/L) levels were in May, and the highest NH₃ (0.43 \pm 0.00 mg/L) level was in June. POM (4.56 \pm 0.03 mg/L) and NO₂ (0.070 \pm 0.00 mg/L) were highest in September, and the highest PO₄ (0.07 \pm 0.00) and S (9.47 \pm 0.01) levels were in August. Mg content was highest in June with the value 2.85 \pm 0.04, and Ca was highest in July with 4.58 \pm 0.01.

DISCUSSION

Stocking density is one of the most important factors affecting growth, yield and survival of cultured species. In general, by increasing stocking density, it will also increase competition for food and space. On the other hand, the effects of stock density on survival are not contentious. While Parsons and Dadswell (1992) reported no direct correlation between the two factors, Karayücel and Karayücel (2000a) claimed the opposite, detecting different survival rates from different stock densities. In the present study, growth characteristics of mussels were expected to be higher for the lower stocking densities compared to the higher densities due to less competition as in Mackie (1984). Our results, however, indicated that there was not an inverse relation between growth and stocking density. This mode of growth pattern may be related to such behavioral characteristics such as proper site selection, food search, and protection against predators. In this study, survival was not affected by stocking density.

It was determined that stocking densities had a significant effect on the live weight and shell length growth of *U. terminalis delicates* for the duration of 6 months (P<0.05). The best growth was achieved in group B (40 ind./m²). Theoretically, individual growth rate of a given species declines with increasing intraspecific density (Beal and Kraus, 2002). However, in the present study, the highest growth did not occurred in the lowest stocking density (Group A).

In this study, the shell length increased from June to the end of August in all groups, however growth performance started to decrease slightly as the temperature drops in the beginning of September. On the other hand, it shell length growth of *U. terminalis delicates* was lower in the colder in the warmer months for all stocking groups. According to Boltovskoy and Cataldo (1999), the golden freshwater mussel (*Limnoperna fortunei*) also grew slower in colder than in warmer months (Boltovskoy and Cataldo, 1999). Also, it was reported that mussel shell growth of *Mytilus edulis* was low in winter (Richardson *et al.*, 1980; Crosby and Dale, 1990). Condition factor is strongly related to temperature and stocking density (Shafee *et al.*, 1998; Karayücel and Karayücel, 2000a). In this study, the highest condition factor was obtained during November in group A (20 ind./ m^2).

Food availability and water quality are considered to be effective on growth during these months. In shallow lakes, the better growth performance has been reported throughout spring and mild winters, under high phytoplankton levels (Chatterji *et al.*, 1984; Spencer and Ellis, 1990; MacIsaac, 1996).On the other hand, specific growth rate began to diminish with temperature increases during July and August. This suggests that growth of *U. terminalis delicates*, depends mainly on food availability hence, an increase in temperature is not a necessity for a higher growth rate.

Oxygen and pH values also have a biological effect on all living organisms (Morris and Corkum, 1999; Karayücel and Karayücel, 2000b). In our study, pH and oxygen values changed with temperature. Some Unionid species are able to live at a pH of 4.7 (the minimum level) and to reproduce and grow at pH values between 5.6-8.3 (McMahon, 1991). pH levels of our study are within the range of 7.78±0.04 to 8.02±0.01, which is in accordance with the intermediate levels for this taxa. Nitrate-nitrogen (NO₃-N) level (1.01-1.43 mg/L) was within the optimum range for living biomass in our study. The low nitrite-nitrogen (NO2-N) value in our study indicated that the lake water has enough oxygen for nitrification. Organic matter concentration in lake water varied between 1.23-3.22 mg/L. This variation might be due to precipitation and runoff. The highest silica value in the lake was 9.47±0.01 in August. The silica level can be low during spring when algae population is the highest, and this level may also show differences depending on season, water flow and precipitation.

Despite the fact that mussel growth is strongly related to annual primary productivity, it is not as related to chlorophyll a concentration (Small and Van Stralen, 1990). In shallow lakes, increased phytoplankton and zooplankton levels do not mean that the season has a direct effect on the chlorophyll a level (Adrian et al., 1999). Our results, confirming to the earlier findings, showed that the mussel growth levels were highest in August, and chlorophyll a levels were highest in April and May. It doesn't necessarily mean that spring has direct effect on chlorophyll a just because phytoplankton and zooplankton rich to maximum concentrations in shallow lakes during spring time (McQuenn et al., 1986; Adrian et al., 1999). However, phosphate (PO₄) content is known to be a limiting factor on Chlorophyll a (Lauritsen, 1986; Spencer and Ellis, 1990; Shapiro, 1995).

In conclusion; the present study was the first attempt investigating optimum stock density for the culture of *U. terminalis delicates*. Since the highest live weight and shell height were obtained from the stocking groups of 40 ind./m², it could be advised to stock the juveniles of *U. terminalis delicates* at a level of 40 ind./m². Also, other unionids should be investigated as a cultivable alternative species.

ACKNOWLEDGEMENTS

We thank the following people who provided laboratory and technical advice and constructive comment on the manuscript: Menderes Şereflişan, İhsan Akyurt Erdal Yılmaz and Kaya Gökçek.

Statement of conflict of interest

Authors have declared no conflict of interest.

REFERENCES

- Adrian, R., Norbert, W., Thomas, H., Sigrid, H. and Rusche, R., 999. Effects of ice duration on plankton succession during spring in a shallow polymictic lake. *Freshw. Biol.*, **41**: 621-632.
- APHA, 1971. American public health association standard methods for the examination of water and wastewater. 13th edition, Washington D.C.
- APHA, 1980. Standard methods for the examination of water and wastewater, 15th ed. American Public Health Association, Washington, D.C.
- Barnhart, M.C., 2006. Buckets of muckets: a compact system for rearing juvenile freshwater mussels *Aquaculture*, **254**: 227–233.
- Beaty, B.B. and Neves, R.J., 1996. Factors influencing the growth and survival of juvenile Villosa iris (Bivalvia: Unionidae) in an artificial stream system. N. Am. J. Aquacul., 15: 483-484.
- Beal, B.F. and Kraus, M.G., 2002. Interactive effects of initial size, stocking density, and type of predator deterrent netting on survival and growth of cultured juveniles of the soft-shell clam, *Mya arenaria* L., in Eastern Maine. *Aquaculture*, **208**: 81-111.
- Bogan, A., 2008. Global diversty of freshwater mussels (Mollusca: Bivalvia) in freshwater. *Hydrobiologia*, **595**: 139-147.
- Boltovskoy, D. and Cataldo, D.H., 1999. Population dynamics of *Limnoperna fortunei*, an invasive fauling mollusc, in the lower Parana River (Argentina). *J. Shellf. Res.*, 14:255-263.
- Chatterji, A., Ansari, Z.A., Ingole, B.S. and Parulekar, A.H., 1984. Growth of the green Mussel, *Perna viridis* L., in a sea water circulating system. *Aquaculture*, **40**:47-55.
- Crosby, M.P. and Gale, L.D., 1990. A review and evaluation of bivalve condition inde methodologies with a suggested standard method. *J. Shellf. Res.*, **9**: 233-237.

- Doroudi, M.S. and Southgate, P.C., 2000. The influence of algal ration and larval density on growth and survival of blacklip pearl oyster *Pinctada margiritifera* (L.) larvae. *Aquacul. Res.*, **42**:178-183.
- Gehrke, C.W., Affsprung, H.E. and Lee, Y.C., 1954. Direct ethylenediaminetetraacetate titration methods for calcium and magnesium. *Anal. Chem.*, **26**:1944.
- Haag, W.R., 2013. The role of fecundity and reproductive effort in defining life-history strategies of North American freshwater mussels. *Biol. Rev.*, 88: 745–766.
- Haag, W. R. and Staton J.L., 2003. Variation in fecundity and other reproductive traits in freshwater mussels. *Freshw. Biol.*, 48:2118-2130
- Hoggarth, M.A., 1999. Descriptions of some of the glochidia of the Unionidae (Mollusca:Bivalvia). *Malacologia*, 41:1-118.
- Karayücel, S. and Karayücel, İ., 2000a. Influence of stock and site on growth, mortality and shell morphology in cultivated blue mussels (*Mytilus edulis* L.) in two Scottish sealochs. *Israeli J. Aquacul.*, **52:** 98-110.
- Karayücel, S. and Karayücel, İ., 2000b. The effect of environmental factors, depth and position on the growth and mortality of raf-cultured blue mussels (*Mytilus edulis* L.) Aquacul. Res., **31**: 893-899.
- Lauritsen, D.D., 1986. Filter feeding, in *Corbicula fluminea* and its effect on seston removal. J. N. Am. Benthol. Soc., 5:165-172.
- Macisaac, H.J., 1996. Potential biotik and abiotic impact of zebra mussel on the inland waters of North America. Am. Zool., 36: 287-299.
- McMahon, R.F., 1991. Mollusca: Bivalvia. ecology and classification of North American freshwater invertebrates (eds. J.H. Thorp and A.P. Covich), Academic Press, San Diego, pp. 315-399.
- Mcquenn, D.J., Post, J.R. and Mills, E.L., 1986.Trophic relationships in freshwater pelagic ecosystems. *Canadian J. Fish. aquat. Sci.*, 43: 1571-1581.
- Monteforte, M. and Garcia-Gasca, A., 1994. Spat collection studies of pearl oysters *Pinctada mazatlonica* and *Pteria sterna* (Bivalvia:Pteridae) in Bay of La Paz, South Baja California, Mexico. *Hydrobiology*, **291**: 21-34.
- Monteforte, M., Bervera, H. and Moroles, S., 1994. Growth and survival of pearl oysters *Pinctada mazatlanica* and *Pteria sterna* in extensive conditions at Bahia da la Paz. South Baja California, Mexico. J. Shellf. Res., **13**: 344-345.
- Monteforte, M. and Morales-Mulia, S., 2000. Growth and survival of the Calafia mother-of-pearl oyster *Pinctada mazatlinica* (Hanley 1856) under different sequences of nursery culture-late culture at Bahia de la Paz, Baja California Sur, Mexico. *Aquacul. Res.*, **31**: 901-915.
- Morris, T.J. and Corkum, L.D., 1999. Unionid growth patterns in rivers of differing riparian vegetation. *Freshw. Biol.*, 42: 59-68.
- Mueller, D. and Patzner, R.A., 1996. Growth and age structure

of the swan mussel *Anodonta cygnea* (L.) at different depths in Lake Mattsee (Salzburg, Austria). *Hydrobiology*, **341**: 65-70.

- Parsons, G.J. and Dadswell, M.J., 1992. Effect of stocking density on growth, production, and survival of the giant scallop, *Placopecta magellanicus*, held in inter mediate suspension culture in Passamaquoddy Bay, New Brunswick. *Aquaculture*, **103**: 291-309.
- Paterson, K.J. and Nell, J.A., 1997. Effect of different growing techniques and substrate types on the growth and survival of the clams *Tapes dorsatus* (Lamarck) and *Katelysia rhytiphora* (Lamy). *Aquacul. Res.*, **28**: 707-715.
- Richardson, C.A., Crisp, D.J., Runham, N.W. and Gruffydd, L.D., 1980. The use of tidal growth bands in the shell of *Cerastoderma edule* to measure seasonal growth rates under cool temperature and sub- arctic conditions. *J. mar. biol. Assoc. U.K.*, 60: 977-989.
- Şereflişan, H., 2014. Gölbaşı Gölü (Hatay) tatlı su midyelerinin ekonomik değer taşıyan özelliklerinin araştırılması. Yunus Araştırma Bül., 3:43-49.
- Shafee, M., Berraho, A. and Raif, M., 1998. Culture of Carpet-Shell clam, *Ruditapes decussatus* (L.) on theAtlantic coast of Morocco. J. Aquacult. Trop., 13: 17-36.
- Shapiro, J., 1995. Lake restoration by biomanipulation-a personal view. *Environ. Rev.*, 3: 83-93.
- Simmons, K.B., Shaffer, V.L., Thacker, S.G. and Luckenbach, M.W., 1995. Hatchery production of eastern oyster, *Crassostrea virginica*, seed: A biological and economic investigation of larval culture techniques including preliminary observations of optimal stocking density and substrate size in a hatchery setting system. *Milfard*

Aquacul., Sem., 14: 247-255.

- Small, A.C. and Van Stralen M.R., 1990.Average annual growth and condition of mussels as a function of food source. *Hydrobiology*, **195**: 179-188.
- Southgate, P.C. and Beer, A.C., 1997. Hatchery and early nursery culture of the blacklip pearl oyster (*Pinctada* margiritifera, L.). J. Shellfish Res., 16: 561-567.
- Spencer, C.N. and Ellis, B.K., 1990. Co-limitation by phosphorus and nitrogen, and effects of zooplankton mortality on phytoplankton in Flathead Lake Montana, U.S.A. Proceedings of the international society for theoretical and Applied. *Limnology*, 24: 206-209.
- Stanczykowska, A. and Levandowski, K.E., 1995. Individual growth of the freshwater mussel *Dreissena polymorpha* (Pall.) in Mikołajskie Lake; estimates in situ. *Ekol Pol*, 43: 267-276.
- Strayer, D. L., Downing, J.A., Haag, W.R., King, T.L., Layzer, J.B., Newton, T.J. and Nichols, S.J., 2004. Changing perspectives on pearly mussels, North America's most imperiled animals. *BioScience*, 54:429-439.
- Şereflişan, H., Çek, Ş. and Şereflişan, M., 2013. The reproductive cycle of *Potomida littoralis* (Cuvier, 1798) (Bivalvia: Unionidae) in Lake Gölbaşi, Turkey. *Pakistan J. Zool.*, **45**:1311-1319.
- Taylor, J.J., Southgate, P.C. and Rose, R.A., 1998. Effect of mesh covers on growth and survival of silver- lip pearl oyster (*Pinctada maxima*, Jameson) spat. *Aquaculture*, 135: 41-49.
- Zar, J.H., 1999. *Biostatistical analysis*. Prentice Hall, New Jersey, pp. 663.